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Mini-symposium Update on primary sclerosing cholangitis

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ABSTRACT

Early studies in primary sclerosing cholangitis (PSC) were concerned with disease characterization, and were followed by epidemiological studies of PSC and clinical subsets of PSC as well as a large number of treatment trials. Recently, the molecular pathogenesis and the practical handling of the patients have received increasing attention. In the present review we aim to give an update on the pathogenesis of PSC and cholangiocarcinoma in PSC, as well as to discuss the current opinion on diagnosis and treatment of PSC in light of the recent European Association for the Study of the Liver and the American Association for the Study of Liver Diseases practice guidelines.

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1. Introduction

Primary sclerosing cholangitis (PSC) is a chronic inflammatory condition leading to fibrotic strictures and dilatations of the bile ducts and in most cases liver cirrhosis [1,2]. Any part of the biliary tract including the gallbladder may be affected. However, the distribution of the affection is typically not uniform, leading to sampling variability on liver histology due to regional inflammation and cholestasis [3]. Dominant strictures of the common or main hepatic ducts develop in approximately half of the patients and predispose for recurrent episodes of bacterial cholangitis [4–6]. PSC should be considered a progressive condition [7], but disease course is highly variable from patient to patient and predicting outcome at the individual level is not possible [8]. The role of medical treatment in PSC is unclear and most patients reach the combined endpoint of death or liver transplantation over a period of 12–17 years following diagnosis [9–11].

Epidemiological studies have found prevalence rates of PSC in Northern European descendants of approximately 10/100.000 [12–14]. In Southern Europe and Asia, the reported numbers are 10–100-fold lower [15,16]. Most PSC patients are relatively young, with a median age at onset of 30–40 years, ranging from children below the age of 10 to elderly individuals 70–80 years old. In contrast to most autoimmune conditions, approximately 2/3 of the PSC patients are male.

An important feature of PSC is the variable presence of comorbidities. Most common is inflammatory bowel disease (IBD), which is reported in the range of 62–83% in Northern European descendants [13]. In Southern Europe and India the IBD frequency is approximately 50% [15,17,18], and in Singapore and Japan even lower (20-37%) [16,19,20]. According to standard criteria, the IBD in PSC in about 80-90% of the cases is compatible with ulcerative colitis (UC) whereas the remainder of the cases are diagnosed with Crohn's disease or IBD unclassified [21]. A variety of other autoimmune diseases have also been reported at an increased frequency in PSC [22], and approximately 6–9% of adult PSC patients have features of autoimmune hepatitis [23-25]. In the range of 3.3-36.4% of the PSC patients will develop cholangiocarcinoma [17,19,26-28], with numbers varying based on geography (lower frequencies in Southern Europe and Asia) and the population studied (higher frequencies in liver transplant center series). Importantly, there is also an increased risk of colonic cancer and gallbladder cancer among the patients [29,30]. It is presently not known whether this heterogeneity means that PSC is a "mixed bag" of multiple conditions still to be defined, or only represents variable disease behaviour of an otherwise homogeneous condition.

As evident from Table 1, years separate landmark discoveries in PSC. In the present review we aim to give an update on the literature on the pathogenesis of PSC and cholangiocarcinoma in PSC, as well as the current opinion on diagnosis and handling of PSC that has been put forward by the practice guidelines recently published by the European Association for the Study of the Liver (EASL) and American Association for the Study of Liver Diseases (AASLD).

2. Pathogenesis of PSC

The etiology and pathogenesis of PSC are not known. By the time diagnosis of PSC can be made by cholangiography, there is already extensive scarring and strictures of the biliary tree. The obstruction of bile flow leads to secondary inflammation and apoptosis

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Table 1

Literature-based history of primary sclerosing cholangitis (PSC). The list is not exhaustive and is intended to illustrate the direction of the field. Early studies were concerned with disease characterization, and were followed by epidemiological studies of PSC and clinical subsets of PSC as well as a large number of unsuccessful treatment trials. Recently, practical handling of the patients and the molecular pathogenesis have received increasing attention and will be the emphasis of this update on PSC.

Year	Landmark finding	Reference
1867	First description of PSC.	[208]
1965	Review of 25 case reports from 1954 to 1964	[209]
	revealed a link between PSC and ulcerative	
	colitis and other autoimmune diseases.	
1966	First single-center case series (42 PSC patients)	[210]
1070	described clinical features of PSC.	[011]
1979	Autoreactivity toward billary antigens in PSC	[211]
1980	Three case series (93 PSC natients in total)	[2 212 21
1500	defined the clinical biochemical radiological	[2,212,21
	and histological features of PSC.	
1982	Association between HLA variants and risk of	[35]
	PSC recognized.	
1983	Liver transplantation established as treatment	[214]
	for end-stage PSC.	
1985	Link between PSC and cholangiocarcinoma	[215]
1095	established.	[216]
1965	Recurrent PSC after liver transplantation	[210]
1500	recognized	[217]
1991	Characteristic features of inflammatory bowel	[21]
	disease in PSC defined.	
1991	Intestinal leakage of bacteria in rats leads to	[102]
	sclerosing cholangitis.	
1992	PSC found to represent a risk factor for colonic	[218]
1002	dysplasia.	[175]
1992	First randomized placebo-controlled trial of	[1/5]
	improvement in biochemistry, but no	
	significant effect on liver transplantation-free	
	survival.	
1992	Features of autoimmune hepatitis in a subset	[135,137]
	of patients with PSC recognized.	
1993	Bile acid toxicity in <i>mdr2</i> knockout mice leads	[77]
1000	to sclerosing cholangitis.	[10]
1998	First epidemiological study of PSC documented	[12]
	A prevalence of approximately 1/10,000 m	
2001	Aberrant homing of intestinally activated	[66]
2001	lymphocytes to liver recognized.	[00]
2005	Increased risk of PSC among relatives	[219]
	demonstrated, underscoring the importance of	
	genetic risk factors.	
2006	Elevated levels of IgG4 in a subset of patients	[130]
2000	with PSC recognized.	[170]
2009	ursedeevychelic increased risk of death or liver	[1/8]
	transplantation	
2009	European Association for the Study of the Liver	[123]
	practice guidelines.	
2009	Genome-wide association study determined	[34]
	the genetic architecture of PSC.	
2010	American Association for the Study of Liver	[124]
	Diseases guidelines.	

[31], and it is thus impossible to determine whether observations at the cellular and molecular level are of primary importance in PSC pathogenesis or only secondary to the ongoing disease processes. Since the biliary tract exhibits a limited repertoire for reacting to any insult, a variety of pathogenetic mechanisms may give rise to a clinical presentation of sclerosing cholangitis [32]. A relevant question is thus whether a single pathogenetic mechanism could be expected in the remaining patients when patients with known causes for sclerosing cholangitis have been excluded. The heterogeneity of the PSC population makes this unlikely. This also means that rather than considering the various hypotheses that have been

Table 2

Leading hypotheses on the pathogenesis of primary sclerosing cholangitis (PSC). Rather than concluding each hypothesis as "true" or "false", they should be considered descriptive of possible components of the disease process in PSC (MAdCAM-1; mucosal addressin cellular adhesion molecule 1, CCL25; chemokine ligand 25, CCR9; chemokine receptor 9, ABCB4; ATP-binding cassette sub-family B member 4, SXR; steroid and xenobiotic recentor. PXR: pregnane X receptor).

Pathogenetic mechanism	Experimental and epidemiological support
The "aberrant homing" hypothesis	MAdCAM-1expression on sinusoidal endothelium during PSC [65,66]. Production of CCL25 in PSC and increased numbers of T-cell expressing CCR9 [67].
The "autoimmunity" hypothesis	Presence of autoantibodies [220]. Preferential usage of particular T-cell receptor gene segments of hepatic T-cell [71]. Strong HLA associations [34]. Autoreactivity in rat model of PSC [221]. Presence of features of autoimmune hepatitis and other autoimmune diseases in some patients [22,23,25,138–144,222].
The "toxic bile" hypothesis	Sclerosing cholangitis in <i>abcb4</i> –/– mice [77,79,80]. Cholestatic liver disease with portal fibrosis, in adults with <i>ABCB4</i> mutations [84,85]. Modifier effects from <i>ABCB4</i> and <i>SXR/PXR</i> gene polymorphisms [86,87]. Development of PSC-like changes in cystic fibrosis [223]. Improvement of hepatic biochemistries under treatment with ursodeoxycholic acid [187].
The "leaky gut" hypothesis	Bacterial translocation in rats induces PSC-like changes [99–103]. Lipopolysaccharide activates toll-like receptors on biliary epithelial cells [107]. Improvement of hepatic biochemistries under treatment with metronidazol [108]. Increased frequency of innate immune cells in PSC livers [58–60].

put forward on the pathogenesis of PSC as "true" or "false" (Table 2), each of the hypotheses could be considered potentially relevant, in some patients, at various stages of PSC.

2.1. Genetics of PSC

The largest study of heritability in PSC has shown that siblings of patients are 9–39 times more likely to develop PSC than the overall population [33]. In the same study, siblings of PSC patients were also found to have an 8-fold increased risk of developing UC even without liver disease, meaning that shared genetic risk factors between PSC and UC are likely to exist. The outcome of a recent genome-wide association study in PSC is schematically shown in Fig. 1 [34]. The strongest associations were detected in the HLA complex on chromosome *6p21*, and weaker associations were found at three genetic loci that had previously been implicated in the susceptibility to IBD (chromosome *3p21*, chromosome *2q35* and the *GPC5/GPC6* region on chromosome *13q31*).

An HLA association in PSC was detected as early as 1982 (Table 1) [35], and the relative importance and PSC specificity of findings within this region clearly warrant priority in further research [34,36]. Probably both the HLA class I and HLA class II genes contribute to the associations. However, since the HLA genes are closely



Fig. 1. The genetic architecture of primary sclerosing cholangitis (PSC). The strong HLA associations represent the only disease specific genetic finding in PSC so far. The remainder of the disease genes have also showed association in the inflammatory bowel diseases (IBD) ulcerative colitis (UC) and Crohn's disease (CD). IL10; interleukin-10, HLA; the human leukocyte antigen complex on chromosome *6p21*, TGR5; takeda G-protein coupled bile acid receptor 5, IL23R; interleukin 23 receptor, IL12B; interleukin 12B, NKX2-3, NK2 transcription factor related locus 3, PTPN2; protein tyrosine phosphatase, non-receptor type 2, CCNY; cyclin Y, STAT3; signal transducer and activator of transcription 3, GPC5/6; glypican 5 and 6, MST1; macrophage-stimulating 1, NOD2; nucleotide-binding oligomerization domain containing 2, IRGM; immunity-related GTPase family M, ATG16L1; ATG16 autophagy related 16-like 1.

linked, it has so far not been possible to conclusively dissect exactly which of the genes that are most important. For the HLA class I genes, two recent studies from Norway and Italy point to the possibility that HLA-C and possibly HLA-B variants with a particularly strong inhibitory influence on T-lymphocytes and natural killer cells via so-called killer immunoglobulin-like receptors may protect against PSC [37,38]. These HLA-C and B variants have low frequencies in Northern Europe where PSC is most prevalent [39]. Several of the HLA class II variants are linked with these HLA-C and HLA-B variants, but the increased risk associated with DRB1*1301 and the decreased risk associated with DRB1*0701 and various DRB1*04 and DRB1*11 variants seem to be due to other mechanisms [37,38,40–42]. One possible explanation is obviously the ability of these or closely linked HLA class II variants to present PSC relevant antigens to the T-lymphocytes.

The finding that only two out of 15 established susceptibility loci for UC also confer risk for PSC corroborates clinical observations that IBD in PSC may represent a distinct entity, different from UC in many aspects ("PSC-IBD") [38,43]. In most patients with IBD and PSC, inflammation is mild with a slight right-sided predominance [21,44,45]. The IBD in PSC is almost always a pancolitis, even when classified as Crohn's disease, however, often with rectal sparing and subtle inflammatory changes in the ileum ("backwash ileitis") [43,46]. What role genetic variants the two UC susceptibility loci may play in the biliary tract is not clear. At chromosome 3p21, the problem of linked genes is a similar problem as in the HLA. One possible candidate is the macrophage-stimulating 1 (MST1) gene, which encodes a circulating protein which inhibits macrophages during inflammation [47]. The PSC-, CD- and UC-associated MST1 variant may impair binding of MST1 to its receptor and thus lead to deficiency of this negative feedback system [48].

At chromosome 2q35, the bile acid receptor TGR5 gene is the most plausible disease gene [49,50]. Like MST1, TGR5 may inhibit macrophages, and the activation of TGR5 by bile acids means that this could prevent excessive inflammation during cholestasis [49]. Furthermore, TGR5 may activate the cystic fibrosis transmembrane conductance regulator (CFTR) in the biliary epithelium [50]. CFTR plays an important role in the protection of the biliary epithelium by means of transportation of chloride ions to bile. These chloride ions are exchanged for bicarbonate by the anionic exchanger 2 (AE2) molecule, and the bicarbonate may constitute one of the mechanisms by which the biliary epithelium is protected against toxic effects from bile [51]. Interestingly, concurrent colitis and CFTR deficiency has been shown to induce PSC-like disease in mice [52]. So far there is no consensus as to a role for CFTR mutations in PSC [53–57], but the possibility should be held open that such variants may be of importance in individual patients.

2.2. Aberrant lymphocyte homing and autoreactivity in PSC

Some studies have reported an increased frequency of natural killer cells in the portal infiltrate of patients with PSC when compared with other liver diseases [58,59], and there is also an increase in Kuppfer cells and peri-sinusoidal macrophages [60]. To what extent this may be related to the genetic findings discussed above is not known. However, the majority of the mononuclear cells in the portal infiltrate in liver biopsies from patients with PSC is Tlymphocytes [61,62]. The "aberrant homing" hypothesis (Table 2) proposes that lymphocytes activated in the gut may be responsible for biliary inflammation in PSC [63]. Normal intestinal homing of T-lymphocytes is ensured by the integrin alpha 4/integrin beta 7 ($\alpha 4\beta 7$) receptor on the lymphocytes and the corresponding mucosal addressin cellular adhesion molecule 1 (MAdCAM-1) ligand on the intestinal endothelial cells [63,64]. In PSC and other inflammatory liver diseases, MAdCAM-1 is expressed on portal vein- and sinusoidal endothelium [65,66], and may thus function to recruit intestinally activated T-lymphocytes to the liver [63,64]. Supportive evidence for the phenomenon comes from the observation of an increased production of the chemokine ligand 25 (CCL25) in PSC [67], which binds the chemokine receptor 9 (CCR9) on memory T-lymphocytes. An open question of the hypothesis is whether observations are specific to PSC or merely a general feature of hepatic inflammation.

The strong HLA association in PSC detected in the genome-wide association study is typical for an autoimmune condition. The high frequency of other autoimmune diseases as well as features of autoimmune hepatitis in PSC further support the presence of an autoimmune component in the pathogenesis. Antibodies against biliary as well as colonic epithelial cells have been reported [68-70], and the preferential usage of particular T-cell receptor gene segments of hepatic T-cells also suggests that tissue-specific antigens of relevance to PSC pathogenesis may exist [71]. The most prevalent autoantibody in PSC (up to 94% of the patients) is a particular type of perinuclear anti-neutrophil cytoplasmatic antibody (pANCA) [72–74]. This antibody is also observed in UC and type 1 autoimmune hepatitis [73,75], and suggests that common pathogenetic mechanisms for these conditions are likely to exist. Recently, tubulin beta 5 (TBB-5) was identified as the main pANCA antigen [76], and antibodies against TBB-5 were found to cross-react with the bacterial FtsZ protein, meaning that an immune response against intestinal bacteria may be the cause of pANCA. Interestingly, anti-FtsZ antibodies were common among healthy controls, whereas the presence of both anti-TBB-5 and anti-FtsZ was predominantly found in autoimmune hepatitis and PSC. The pathogenetic importance of pANCA and other autoantibodies in PSC is not known and should be assessed in parallel with further studies on the HLA association in PSC.

2.3. Bile acid toxicity in PSC

Several studies have elaborated on the "toxic bile" hypothesis in PSC pathogenesis (Table 2), based on the findings of PSC-like changes in mice devoid of the phospholipid transporter "multidrug resistance protein 2" (mdr2), called MDR3 and ABCB4 in humans [77,78]. In these mice, bile acid toxicity leads to extensive fibrosis of the bile ducts mimicking intrahepatic PSC in humans [79-82]. Whereas defects in the human ABCB4 gene leads to progressive familial intrahepatic cholestasis type 3 (PFIC3) [83], no associations between ABCB4 variants and PSC have so far been reported. However, single cases of ABCB4-related disease may be found among PSC patients without IBD, as highlighted by the findings of "mild" ABCB4 mutations in adult patients with unexplained cholestasis and histological findings of portal fibrosis and minor ductular changes in two recent studies [84,85]. Furthermore, similar mutations may alter disease severity ("modifier effects"), as demonstrated by the findings of a more severe disease course in both PSC and primary biliary cirrhosis (PBC) patients with particular ABCB4 variants [86,87].

Bile acids that accumulate in affected segments of the liver in PSC are toxic to the hepatocytes, and are likely to contribute to the development of liver cirrhosis in PSC [88]. The hepatocyte initiates a variety of detoxification mechanisms in cholestasis, including hydroxylation and conjugation to make bile acids more water soluble and induction of bile acid transporters on the sinusoidal membrane leading to export of metabolized bile acids to the systemic circulation for renal elimination [89]. One of the major determinants of the expression of the genes involved in these processes is the steroid and xenobiotic receptor (SXR; also called pregnane X receptor, PXR) [90,91]. PSC patients with particular genetic variants of SXR have been shown to suffer a more severe disease course, in line with the importance of the protective effects from SXR in animal models of cholestasis [89,92-96]. Since variants of SXR have recently also been shown to influence disease severity and liver damage in PBC and non-alcoholic fatty liver disease [97,98], the mechanisms involved could be of general importance for liver cell injury.

2.4. The "leaky gut" hypothesis

Studies in a rat model with intestinal bacterial overgrowth performed almost 20 years ago suggested that innate immune responses to bacterial products may initiate a PSC-like pathogenesis [99–103]. The findings warrant renewed interest for several reasons. Since the inflammation in these models was mediated by TNF α , it is of great interest to note increased levels of this pro-inflammatory cytokine also in human PSC as compared with healthy controls and other chronic cholestatic liver diseases [104–106]. Along with macrophages, both natural killer cells and T-lymphocytes can produce TNF α . Since several of the genetic studies in PSC implicate a role of innate immune responses, including macrophages and natural killer cells, the relationship between these genetic findings and translocation of bacterial components in an inflamed colon needs to be further explored.

Interestingly, toll-like receptor 4 (TLR-4) and TLR-9 expression on biliary epithelial cells may be induced by antibodies against these cells [107]. These TLRs recognize bacterial products and viral DNA and activation may aggravate ongoing biliary inflammation upon the exposure to bacterial or viral components in blood. Such components may be derived from bacterial colonization of the biliary tree, or even infections at other mucosal surfaces or flares of IBD. Whether such mechanisms could help explain the fluctuating disease course observed in many PSC patients is not known, but it is interesting to note an improvement of hepatic biochemistries in PSC patients under long-term administration of metronidazol [108]. The presence of an infectious trigger or infectious modifier effects in PSC is thus likely, but further studies are required to determine the type and importance of such factors. Possibly, a diverse spectrum of infectious agents may cause breakdown of immunological tolerance in the bile ducts in genetically susceptible individuals, leading to sustained immunological reactions towards "self-antigens" even after the infectious agents themselves have been cleared. In this regard, it is interesting to note that induction of pANCA seems to be dependent on the intestinal flora or intestinal inflammation [76].

3. Pathogenesis of cholangiocarcinoma in PSC

The mechanisms of carcinogenesis in PSC are poorly understood [109]. Regarding carcinogenic effects from bile acids, they seem to be in part mediated via DNA damage caused by reactive oxygen intermediates [110]. Defects of the DNA repair machinery have been detected in some patients with cholangiocarcinoma and may aggravate such effects [111]. A series of studies by Gores et al. have demonstrated the importance of autocrine effects from IL-6 on cholangiocarcinoma cell immortalization [112]. The effects from IL-6 are mediated via activation of the signal transducer and activator of transcription 3 (STAT3) protein which leads to the upregulation of several molecules involved in cancer cell apoptosis and proliferation [113]. Normally, the STAT3 activation is restricted via negative feedback by Suppressor of cytokine signaling 3 (SOCS3). In cholangiocarcinoma, however, this negative feedback is defective due to inactivation of the promoter region of the SOCS3 gene by methylation of the DNA [114]. As recently reviewed elsewhere, similar mechanisms (called epigenetic alterations) may also be of more widespread importance in cholangiocarcinoma development [115].

As part of the ongoing inflammation, and in response to the altered properties of the dysplastic cholangiocytes, various immune cells play important roles in cholangiocarcinogenesis. Some of these cells may promote tumor growth and metastasis (e.g. macrophages) [116], whereas others are trying to kill abnormal cholangiocytes (e.g. natural killer cells) [117]. The importance of this balance was recently demonstrated by the finding of an association between genetic variants of the natural killer cell receptor NKG2D and cholangiocarcinoma [118]. In line with findings in other types of cancers [119], individuals who carried NKG2D variants that lead to less efficient killing by natural killer cells were more prone to develop cholangiocarcinoma than individuals with normal NK cells. Very characteristically, approximately 40-50% of the cholangiocarcinomas are diagnosed during the first year following the diagnosis of PSC [10,120,121]. This contrasts the situation of many other inflammation-related cancers (e.g. colonic carcinoma in UC), where the risk gradually increases over the years. In PSC, less than 10% of the patients develop cholangiocarcinoma more than 10 years after the diagnosis [120]. Based on all these observations it could be speculated that "PSC with cholangiocarcinoma" may represent a distinct clinical subset, and that the risk does not apply to all patients.

4. Diagnostic challenges in PSC

The diagnosis of PSC is made by cholangiography and the exclusion of secondary etiologies. There are no specific symptoms or clinical, biochemical or histological findings. There are important challenges regarding (1) the definition of "outlier" PSC variants (small duct PSC, autoimmune hepatitis-like PSC and IgG4-associated cholangitis), (2) PSC in children and (3) early diagnosis of biliary and colonic malignancies. Regarding the cholangiogram, the typical findings in PSC involve multifocal strictures and dilatations

Table 3

Summary points of the EASL and AASLD practice guidelines on primary sclerosing cholangitis (PSC) [123,124]. MRC; magnetic resonance cholangiography, ERC; endoscopic retrograde cholangiography, IBD; inflammatory bowel disease, UDCA; ursodeoxycholic acid.

Practice point	EASL guidelines	AASLD guidelines
Cholangiography	MRC recommended as initial investigation.	MRC recommended as initial investigation.
	ERC if indicated.	ERC if indicated.
Liver biopsy—adults	Only in patients with normal cholangiography	Only in patients with normal cholangiography
	or disproportionally elevated serum	or disproportionally elevated serum
	transaminases.	transaminases.
Liver biopsy—children	Recommended.	Recommended.
Antibiotic prophylaxis during ERC	Recommended.	No recommendation point.
Long-term antibiotic treatment	No recommendation point.	Recommended in patients with recurrent
		attacks of acute cholangitis.
Endoscopic treatment	Balloon dilatation with or without stenting.	Balloon dilatation with or without stenting.
UDCA treatment in PSC	No specific recommendation made.	Not recommended.
UDCA chemoprevention	In patients with longstanding IBD and family	Not recommended.
	history of colorectal malignancies.	
Treatment of PSC with features of autoimmune hepatitis	UDCA and immunosuppression recommended.	Corticosteroids and other immunosuppressive agents recommended.
Treatment of IgG4-associated sclerosing cholangitis	Corticosteroids and/or azathioprine.	No specific treatment recommendation.
Liver transplantation	Treatment of choice in cirrhotic patients and	Treatment of choice in cirrhotic patients and
-	should be considered in refractory bacterial	should be considered in refractory bacterial
	cholangitis.	cholangitis.
Surveillance colonoscopy	Every 1–2 years in IBD.	Every 1–2 years in IBD.
Surveillance ultrasound	Annually.	Annually.
Cholangiocarcinoma surveillance	No recommendation made.	No recommendation made.

of both the intra- and the extra-hepatic bile ducts [122]. Changes of the extra-hepatic bile ducts only are rare, whereas isolated changes of the intrahepatic bile ducts have been reported at a frequency of 20–28% of the patients [4,10,27,122]. Whereas both the EASL and the AASLD guidelines now recommend MRC rather than ERC as the first investigation in patients where PSC is suspected (Table 3) [123,124], the quality of the investigation needs careful assessment and ERC should be performed in inconclusive cases. There have been concerns as to the sensitivity of MRC in detecting subtle intrahepatic changes [125], and some authors also claim that MRC is inferior to ERC in the detailed characterization of extra-hepatic biliary changes and dominant strictures [126,127].

4.1. Diagnosis of variant forms of PSC

As the etiologies of what we today denominate "primary" in relation to sclerosing cholangitis are unravelled, the list of differential diagnoses in terms of secondary sclerosing cholangitis is likely to grow. The ultimate outcome of the ongoing research on the pathogenesis of PSC could be the replacement of the PSC diagnosis by a distinct set of etiology-based diagnoses. Whether small duct PSC, PSC showing features of autoimmune hepatitis (also called "PSC-autoimmune hepatitis overlap syndrome" and "autoimmune sclerosing cholangitis") and IgG4-associated sclerosing cholangitis might represent distinct diagnoses has to be determined.

In the EASL and AASLD practice guidelines, IgG4-associated sclerosing cholangitis has now been placed on the list of etiologies for secondary sclerosing cholangitis. Interestingly, some of the IgG4associated sclerosing cholangitis patients may have IBD, and like in regular PSC, there is a distinct male predominance [128,129]. Elevated levels of IgG4 (>135 mg/dl) are specific for the diagnosis. However, sensitivity may be lower than previously reported (71–82%) [128], and repeated measurements may be required to establish the diagnosis [129]. Whereas elevated levels of IgG4 have been reported in 7–9% of PSC patients [19,130], positive immunostaining for IgG4 was recently found in 23% of liver explant specimens from patients with PSC [128]. These data indicate that work remains in delineating IgG4-associated sclerosing cholangitis from regular PSC. The importance of establishing the diagnosis, however, is the excellent response to corticosteroid treatment.

The EASL and AASLD guidelines agree on the need for liver biopsy to exclude small duct PSC in patients with clinical and biochemical features of PSC but with a normal cholangiogram. Whereas the presence of IBD is no longer mandatory for the diagnosis of small duct PSC in either of the guidelines, liver biopsy interpretation relies on IBD status. In patients without IBD, typical changes suggestive of PSC are required. In patients with concurrent IBD, the histological changes should at least be compatible with PSC, but not necessarily typical or specific for regular PSC. As highlighted by the EASL guidelines, abandoning the IBD criterion for small duct PSC may lead to misclassification of patients with "mild" forms of hereditary cholestatic syndromes (e.g. ABCB4disease) as small duct PSC [84,85,123]. In familial cases without IBD, mutational analysis may thus be advisable. Transition to large duct PSC may occur, and cholangiography should be repeated on clinical detoriation [131–134]. Typically, however, small duct PSC runs a quiescent course, and long-term survival is significantly better than for regular PSC [131,132]. Cholangiocarcinoma does not seem to occur in small duct PSC patients unless transition into regular PSC has occurred [131].

The presence of biochemical, serological and histological features of autoimmune hepatitis in a subset of PSC patients has long been recognized [135-137]. The nomenclature for this group of patients is not clear, but both the EASL and AASLD guidelines have adopted the term PSC-autoimmune hepatitis overlap syndrome. A liver biopsy should be considered in patients with an autoimmune hepatitis-like autoantibody profile (high levels of anti-nuclear antibodies [ANA] and anti-smooth muscle antibodies [anti-SMA]) or relatively high IgG and serum aminotransferase levels to assess features of autoimmune hepatitis [123,138]. By the application of recent scoring criteria for autoimmune hepatitis, approximately 6-9% of adult PSC patients seem to have features of autoimmune hepatitis [23-25], and in pediatric PSC patients the frequency is even higher [139–144]. The importance of detecting the presence of autoimmune hepatitis-like features in PSC are reports suggesting a more benign disease course associated with immunosuppressive therapy in this group of patients [138,145,146].

4.2. Diagnosis of PSC in children

The main challenges in the diagnosis of PSC in children is the slightly different spectrum of causes of secondary sclerosing cholangitis to be excluded and the high frequency of autoimmune hepatitis-like features. In contrast to adult PSC, where a liver biopsy is not recommended, the AASLD guidelines therefore recommend liver biopsy to be performed in all children where PSC is suspected [124]. Some of the differential diagnoses in childhood PSC are listed in the EASL practice guidelines [123], and cover a variety of conditions (in particular cystic fibrosis, α1-antitrypsin deficiency, immunodeficiency syndromes, Langerhans cell histiocytosis, idiopathic neonatal sclerosing cholangitis, biliary atresia, congenital bile duct abnormalities and progressive familial intrahepatic cholestasis type 3). The subgroup of pediatric PSC patients with features of type 1 autoimmune hepatitis has been denominated autoimmune sclerosing cholangitis [144], but further studies are needed to clarify if this demarcation versus regular PSC is justified. A particular point regarding pediatric PSC is the low risk of cholangiocarcinoma and colorectal cancer, meaning that in patients with PSC and IBD below the age of 16 years neither surveillance for cholangiocarcinoma nor annual colonoscopy is recommended [124].

5. Diagnostic challenges in cholangiocarcinoma in PSC

An early diagnosis of cholangiocarcinoma in PSC is difficult to obtain. Metastatic disease has been reported at diagnosis in approximately half of the PSC patients with cholangiocarcinoma [121,147,148], and in approximately 30–40% of the cases the diagnosis was settled in conjunction with liver transplantation for PSC [120,149]. These problems are reflected by the lack of surveillance strategies for cholangiocarcinoma in both the EASL and the AASLD guidelines [123,124]. However, since the increased risk of biliary malignancies in PSC also pertains to the gallbladder [29], annual abdominal ultrasonography to detect gallbladder polyps is recommended. Furthermore, both the EASL and AASLD guidelines advice for colonoscopy with biopsies in all patients with PSC and IBD to allow for the early detection of colorectal cancer.

A problem in diagnosing cholangiocarcinoma in PSC is that the growth is often longitudinal with subtle perineural and perivascular invasion [150], meaning that in many of the patients the tumor cannot be detected by radiology. Contrast-enhanced magnetic resonance (MR) imaging with MRC is considered the imaging method of choice if a cholangiocarcinoma is suspected [151-153]. In case of pathological findings, CT typically supplements with information on lymph node enlargement and the liver parenchyma [152,154] and has been reported superior to MR imaging in terms of defining extra-hepatic growth and vascular encasement [155,156]. Positron emission tomography (PET) scanning is useful for detecting solid metastases [157], but has in recent studies proven unreliable for detecting peritoneal carcinomatosis or biliary lesions not visible on MRI/CT [158-160]. By cholangioscopy, a sensitivity of 92% and specificity of 93% in the diagnosis of malignant strictures in PSC were recently reported [161], as compared with 66% and 51%, respectively, for ERC alone. The application of intraductal ultrasound correspondingly increased the sensitivity from 63% to 88% and specificity from 53% to 91%, respectively, when compared to ERC alone [162]. As emphasized by both the EASL and AASLD guidelines, further studies on the utility of these two latter methods in the diagnosis of cholangiocarcinoma in PSC are needed.

An extensive evaluation of the tumor marker carbohydrate antigen (CA) 19-9, imaging and brush cytology-based techniques in cholangiocarcinoma in PSC was recently published [163]. As also shown in previous studies [164–166], digital image analysis (DIA) and fluorescent in situ hybridization (FISH) enhanced the sensitivity obtained by routine brush cytology investigations. Furthermore, the authors propose that repeated CA 19-9 measurements along with imaging may be useful for the screening and surveillance of cholangiocarcinoma in PSC. By application of the proposed guidelines, almost two-thirds of the cholangiocarcinomas were detected at an early stage where potentially curative liver transplantation protocols were still applicable. More refined methods for early cholangiocarcinoma detection in PSC are also clearly needed. Several "-omics" are presently at work to identify more specific tumor markers, that may be derived from cancer specific protein profiles (proteomics) [167], mutations (genomics), altered regulation of gene expression (epigenomics) [115,168,169], or small-molecular metabolites (metabolomics) [170]. The first results from some of these studies are encouraging [171,172], but it is not yet clear which method is most relevant and replication of the findings and evaluation as screening tools have not been performed for any of the candidates.

6. Treatment of PSC

The scarcity of PSC patients and the long time until a primary endpoint like death or liver transplantation is reached, means that achieving an adequate study population in randomized, doubleblinded treatment trials in PSC is difficult. Furthermore, patients show a remarkable variability in natural history, in contrast to other liver diseases like primary biliary cirrhosis which follows a relatively predictable course [8]. This means that selection bias is a considerable source of error and that the application of surrogate markers for disease progression (e.g. prognostic indices) is likely to yield an imprecise picture of treatment effects.

While the first three studies on ursodeoxycholic acid were able to document an improvement of both hepatic biochemistries and histological parameters [173-175], no improvement in transplantfree survival was observed. Which components of the disease process that is altered by ursodeoxycholic acid in PSC (Table 2) and could explain these findings remain to be defined. The three largest studies to date were performed in the US and Scandinavia and utilized different doses of ursodeoxycholic acid (13–15 mg/(kg day) [176], 17–23 mg/(kg day) [177], and 28–30 mg/(kg day) [178]). In none of these studies significant effects on risk of liver transplantation or death were detected, as also evident from the recent meta-analysis of ursodeoxycholic acid trials in PSC [179]. Furthermore, contrary to suggestions made by pilot studies of high-dose ursodeoxycholic acid in PSC [180,181], the highest dose regimen (28-30 mg/(kg day)) was significantly associated with an increased risk of liver transplantation or death compared with placebo. The explanation for this is not clear, but one speculation made was that higher doses could result in increased colonic conversion of unabsorbed ursodeoxycholic acid into the toxic metabolite lithocholic acid [178]. Based on the sum of these trials, routine prescription of ursodeoxycholic acid in patients with PSC is not recommended in the EASL and AASLD guidelines.

Several studies have addressed the question of whether ursodeoxycholic acid could protect against the development of cholangiocarcinoma or colorectal cancer in PSC. While there are only anecdotal reports regarding a protective effect against cholangiocarcinoma [182,183], two small retrospective series and one small prospective study have suggested a decreased risk of colonic dysplasia associated with ursodeoxycholic acid use [184-186]. However, since none of the large prospective high-dose ursodeoxycholic acid treatment trials have been able to confirm similar effects [177,178], the AASLD guidelines conclude by recommending against prescription of ursodeoxycholic acid as a chemopreventive agent in PSC. The EASL guidelines do not conclude on the issue, and opens up for low-dose ursodeoxycholic acid in PSC patients when additional risk factors are present (e.g. family history of colorectal cancer, previous colorectal neoplasia or longstanding extensive colitis).

A variety of immunosuppressive drugs ranging from corticosteroids to monoclonal antibodies against tumor necrosis factor alpha have been tested in pilot studies as well as randomized double-blind placebo-controlled treatment trials in PSC [187]. None of these agents have proven beneficial and they should not be prescribed to the regular PSC patient. In PSC patients with features of autoimmune hepatitis and IgG4-associated sclerosing cholangitis, corticosteroid treatment with or without adjuvant immunosuppressive therapy is recommended (Table 3). In PSC patients with features of autoimmune hepatitis it is important to recognize that progression to cirrhosis occurs in a majority of the patients despite treatment [188], indicating that some of the pathologic processes may be relatively inert to immunosuppression. In IgG4-associated sclerosing cholangitis, prevention and handling of relapse may represent a considerable challenge [189–191].

Three retrospective studies have noted a significant increase in survival as compared with predicted 3- and 5-year survival rates (according to the Mayo risk score) following endoscopic treatment of dominant stenoses [192–194]. The presence of dominant strictures was also recently reported to associate with poor outcome in PSC [6]. According to both the EASL and AASLD practice guidelines (Table 3), dominant strictures in PSC with significant cholestasis should be treated with balloon dilatation [193,195,196]. Some patients also appear to benefit from short-term stenting [197,198]. No randomized, prospective controlled trials have been performed to assess the efficacy of endoscopic treatment in PSC, and the application is presently performed based on individual assessment of each patient.

In the Nordic countries, PSC is the most important indication for liver transplantation. In the US, PSC is among the five leading indications, and even in low-prevalence countries like Italy and Spain, PSC is among the ten most common indications. Patient survival is excellent, with recent 1 and 5-year survival rates at most centers approaching 90% and 85%, respectively. However, liver transplantation in PSC poses several particular challenges. Disease course is unpredictable, and some patients may require listing for liver transplantation before end-stage liver disease. Listing of PSC patients for liver transplantation on the basis of refractory bacterial cholangitis even in non-cirrhotic patients is now recommended in the EASL and AASLD guidelines (Table 3). The high risk of biliary and colonic malignancies means that thorough pre-transplant evaluation as to the presence of cancer must be performed [199,200].

An increased risk of acute cellular rejection in PSC has been demonstrated in several series [201-203]. Since high frequencies of acute cellular rejection have also been reported in recipients with an underlying autoimmune hepatitis and primary biliary cirrhosis, it is not clear whether PSC patients are particularly at risk or whether the risk is related to autoimmune liver disease in general. Furthermore, there is an increased risk of acute rejection in patients with pre-transplant IBD as compared with patients without IBD [202,204-206], and the risk of chronic rejection also seems to be higher in PSC patients with IBD [202]. Based on all these observations, some authorities recommend an intensified immunosuppressive regimen including life-long corticosteroids following liver transplantation for PSC [199], whereas others cautiously prefer a regular regimen based on a study showing an association between aggressive immunosuppression and recurrent PSC [207].

7. Conclusion

Despite the many important discoveries made over the last three decades (Table 1), many important questions related to PSC remain unanswered. Further characterization of the etiology and pathogenesis of PSC is required and should be performed on the basis of genetic risk factors and the pathogenetic knowledge already available (Table 2). Such studies may explain the heterogeneity of the patients and propose targets for novel therapeutics. The development of tools for early diagnosis of PSC in UC is also a priority,

since the fibrotic stage at which the biliary pathology can be diagnosed by cholangiography may prove resistant to medical therapy. Furthermore, sensitive and specific markers for the prediction and diagnosis of cholangiocarcinoma are urgently needed, and novel markers to predict disease behavior in the individual patient would allow for adequate timing of liver transplantation as well as assessment of response to therapy.

Conflict of interest statement

None declared.

References

- Ludwig J, MacCarty RL, LaRusso NF, et al. Intrahepatic cholangiectases and large-duct obliteration in primary sclerosing cholangitis. Hepatology 1986;6:560–8.
- [2] Wiesner RH, LaRusso NF. Clinicopathologic features of the syndrome of primary sclerosing cholangitis. Gastroenterology 1980;79:200–6.
- [3] Olsson R, Hagerstrand I, Broome U, et al. Sampling variability of percutaneous liver biopsy in primary sclerosing cholangitis. J Clin Pathol 1995;48:933–5.
- [4] Tischendorf JJ, Hecker H, Kruger M, et al. Characterization, outcome, and prognosis in 273 patients with primary sclerosing cholangitis: a single center study. Am J Gastroenterol 2007;102:107–14.
- [5] Bjornsson E, Lindqvist-Ottosson J, Asztely M, et al. Dominant strictures in patients with primary sclerosing cholangitis. Am J Gastroenterol 2004;99:502–8.
- [6] Rudolph G, Gotthardt D, Kloters-Plachky P, et al. Influence of dominant bile duct stenoses and biliary infections on outcome in primary sclerosing cholangitis. J Hepatol 2009;51:149–55.
- [7] Porayko MK, LaRusso NF. Wiesner RH Primary sclerosing cholangitis: a progressive disease? Semin Liver Dis 1991;11:18–25.
- [8] Wiesner RH. Liver transplantation for primary sclerosing cholangitis: timing, outcome, impact of inflammatory bowel disease and recurrence of disease. Best Pract Res Clin Gastroenterol 2001;15:667–80.
- [9] Farrant JM, Hayllar KM, Wilkinson ML, et al. Natural history and prognostic variables in primary sclerosing cholangitis. Gastroenterology 1991;100:1710-7.
- [10] Broome U, Olsson R, Loof L, et al. Natural history and prognostic factors in 305 Swedish patients with primary sclerosing cholangitis. Gut 1996;38:610–5.
- [11] Aadland E, Schrumpf E, Fausa O, et al. Primary sclerosing cholangitis: a longterm follow-up study. Scand J Gastroenterol 1987;22:655-64.
- [12] Boberg KM, Aadland E, Jahnsen J, et al. Incidence and prevalence of primary biliary cirrhosis, primary sclerosing cholangitis, and autoimmune hepatitis in a Norwegian population. Scand J Gastroenterol 1998;33:99–103.
- [13] Kingham JG, Kochar N, Gravenor MB. Incidence, clinical patterns, and outcomes of primary sclerosing cholangitis in South Wales, United Kingdom. Gastroenterology 2004;126:1929–30.
- [14] Bambha K, Kim WR, Talwalkar J, et al. Incidence, clinical spectrum, and outcomes of primary sclerosing cholangitis in a United States community. Gastroenterology 2003;125:1364–9.
- [15] Escorsell A, Pares A, Rodes J, et al. Epidemiology of primary sclerosing cholangitis in Spain. J Hepatol 1994;21:787–91.
- [16] Ang TL, Fock KM, Ng TM, et al. Clinical profile of primary sclerosing cholangitis in Singapore. J Gastroenterol Hepatol 2002;17:908–13.
- [17] Okolicsanyi L, Fabris L, Viaggi S, et al. Primary sclerosing cholangitis: clinical presentation, natural history and prognostic variables: an Italian multicentre study. The Italian PSC Study Group. Eur J Gastroenterol Hepatol 1996;8:685–91.
- [18] Kochhar R, Goenka MK, Das K, et al. Primary sclerosing cholangitis: an experience from India. J Gastroenterol Hepatol 1996;11:429–33.
- [19] Takikawa H, Takamori Y, Tanaka A, et al. Analysis of 388 cases of primary sclerosing cholangitis in Japan; Presence of a subgroup without pancreatic involvement in older patients. Hepatol Res 2004;29:153–9.
- [20] Takikawa H, Manabe T. Primary sclerosing cholangitis in Japan–analysis of 192 cases. J Gastroenterol 1997;32:134–7.
- [21] Fausa O, Schrumpf E, Elgjo K. Relationship of inflammatory bowel disease and primary sclerosing cholangitis. Semin Liver Dis 1991;11:31–9.
- [22] Saarinen S, Olerup O, Broome U. Increased frequency of autoimmune diseases in patients with primary sclerosing cholangitis. Am J Gastroenterol 2000;95:3195–9.
- [23] Kaya M, Angulo P, Lindor KD. Overlap of autoimmune hepatitis and primary sclerosing cholangitis: an evaluation of a modified scoring system. J Hepatol 2000;33:537–42.
- [24] Alvarez F, Berg PA, Bianchi FB, et al. International Autoimmune Hepatitis Group Report: review of criteria for diagnosis of autoimmune hepatitis. J Hepatol 1999;31:929–38.
- [25] van Buuren HR, van Hoogstraten HJE, Terkivatan T, et al. High prevalence of autoimmune hepatitis among patients with primary sclerosing cholangitis. J Hepatol 2000;33:543–8.
- [26] Miros M, Kerlin P, Walker N, et al. Predicting cholangiocarcinoma in patients with primary sclerosing cholangitis before transplantation. Gut 1991;32:1369–73.

- [27] Helzberg JH, Petersen JM, Boyer JL. Improved survival with primary sclerosing cholangitis. A review of clinicopathologic features and comparison of symptomatic and asymptomatic patients. Gastroenterology 1987;92: 1869–75.
- [28] Ataseven H, Parlak E, Yuksel I, et al. Primary sclerosing cholangitis in Turkish patients: characteristic features and prognosis. Hepatobiliary Pancreat Dis Int 2009;8:312–5.
- [29] Karlsen TH, Schrumpf E, Boberg KM. Gallbladder polyps in primary sclerosing cholangitis: not so benign. Curr Opin Gastroenterol 2008;24:395–9.
- [30] Soetikno RM, Lin OS, Heidenreich PA, et al. Increased risk of colorectal neoplasia in patients with primary sclerosing cholangitis and ulcerative colitis: a meta-analysis. Gastrointest Endosc 2002;56:48–54.
- [31] Guicciardi ME, Gores GJ. Cholestatic hepatocellular injury: what do we know and how should we proceed. J Hepatol 2005;42:297–300.
- [32] Abdalian R, Heathcote EJ. Sclerosing cholangitis: a focus on secondary causes. Hepatology 2006;44:1063-74.
- [33] Bergquist A, Montgomery SM, Bahmanyar S, et al. Increased risk of primary sclerosing cholangitis and ulcerative colitis in first-degree relatives of patients with primary sclerosing cholangitis. Clin Gastroenterol Hepatol 2008;6:939–43.
- [34] Karlsen TH, Franke A, Melum E, et al. Genome-wide association analysis in primary sclerosing cholangitis. Gastroenterology, doi:10.1053/j.gastro.2009.11.046.
- [35] Schrumpf E, Fausa O, Forre O, et al. HLA antigens and immunoregulatory T cells in ulcerative colitis associated with hepatobiliary disease. Scand J Gastroenterol 1982;17:187–91.
- [36] Karlsen TH, Boberg KM, Vatn M, et al. Different HLA class II associations in ulcerative colitis patients with and without primary sclerosing cholangitis. Genes Immun 2007;8:275–8.
- [37] Karlsen TH, Boberg KM, Olsson M, et al. Particular genetic variants of ligands for natural killer cell receptors may contribute to the HLA associated risk of primary sclerosing cholangitis. J Hepatol 2007;46:899–906.
- [38] Hov JR, Lleo A, Selmi C, et al. Genetic associations in Italian primary sclerosing cholangitis: heterogeneity across Europe defines a critical role for HLA-C. J Hepatol; in press.
- [39] Guinan KJ, Cunningham RT, Meenagh A, et al. Receptor systems controlling natural killer cell function are genetically stratified in Europe. Genes Immun 2010;11:67–78.
- [40] Donaldson PT, Norris S. Evaluation of the role of MHC class II alleles, haplotypes and selected amino acid sequences in primary sclerosing cholangitis. Autoimmunity 2002;35:555–64.
- [41] Wiencke K, Karlsen TH, Boberg KM, et al. Primary sclerosing cholangitis is associated with extended HLA-DR3 and HLA-DR6 haplotypes. Tissue Antigens 2007;69:161–9.
- [42] Spurkland A, Saarinen S, Boberg KM, et al. HLA class II haplotypes in primary sclerosing cholangitis patients from five European populations. Tissue Antigens 1999;53:459–69.
- [43] Loftus Jr EV, Harewood GC, Loftus CG, et al. PSC-IBD: a unique form of inflammatory bowel disease associated with primary sclerosing cholangitis. Gut 2005;54:91–6.
- [44] Oshitani N, Jinno Y, Sawa Y, et al. Does colitis associated with primary sclerosing cholangitis represent an actual subset of ulcerative colitis? Hepatogastroenterology 2003;50:1830–5.
- [45] Joo M, Abreu-e-Lima P, Farraye F, et al. Pathologic features of ulcerative colitis in patients with primary sclerosing cholangitis: a case-control study. Am J Surg Pathol 2009;33:854–62.
- [46] Abdelrazeq AS, Wilson TR, Leitch DL, et al. Ileitis in ulcerative colitis: is it a backwash? Dis Colon Rectum 2005;48:2038–46.
- [47] Zhou YQ, Chen YQ, Fisher JH, et al. Activation of the RON receptor tyrosine kinase by macrophage-stimulating protein inhibits inducible cyclooxygenase-2 expression in murine macrophages. J Biol Chem 2002;277:38104-10.
- [48] Goyette P, Lefebvre C, Ng A, et al. Gene-centric association mapping of chromosome 3p implicates MST1 in IBD pathogenesis. Mucosal Immunol 2008;1:131–8.
- [49] Keitel V, Donner M, Winandy S, et al. Expression and function of the bile acid receptor TGR5 in Kupffer cells. Biochem Biophys Res Commun 2008;372:78–84.
- [50] Keitel V, Cupisti K, Ullmer C, et al. The membrane-bound bile acid receptor TGR5 is localized in the epithelium of human gallbladders. Hepatology 2009;50:861–70.
- [51] Beuers U. Drug insight: mechanisms and sites of action of ursodeoxycholic acid in cholestasis. Nat Clin Pract Gastroenterol Hepatol 2006;3: 318–28.
- [52] Blanco PG, Zaman MM, Junaidi O, et al. Induction of colitis in cftr-/mice results in bile duct injury. Am J Physiol Gastrointest Liver Physiol 2004;287:G491-6.
- [53] Sheth S, Shea JC, Bishop MD, et al. Increased prevalence of CFTR mutations and variants and decreased chloride secretion in primary sclerosing cholangitis. Hum Genet 2003;113:286–92.
- [54] Girodon E, Sternberg D, Chazouilleres O, et al. Cystic fibrosis transmembrane conductance regulator (CFTR) gene defects in patients with primary sclerosing cholangitis. J Hepatol 2002;37:192–7.
- [55] Henckaerts L, Jaspers M, Van Steenbergen W, et al. Cystic fibrosis transmembrane conductance regulator gene polymorphisms in patients with primary sclerosing cholangitis. J Hepatol 2009;50:150–7.

- [56] Pall H, Zielenski J, Jonas MM, et al. Primary sclerosing cholangitis in childhood is associated with abnormalities in cystic fibrosis-mediated chloride channel function. J Pediatr 2007;151:255–9.
- [57] Gallegos-Orozco JF, EY C, Wang N, et al. Lack of association of common cystic fibrosis transmembrane conductance regulator gene mutations with primary sclerosing cholangitis. Am J Gastroenterol 2005;100:874–8.
- [58] Hashimoto E, Lindor KD, Homburger HA, et al. Immunohistochemical characterization of hepatic lymphocytes in primary biliary cirrhosis in comparison with primary sclerosing cholangitis and autoimmune chronic active hepatitis. Mayo Clin Proc 1993;68:1049–55.
- [59] Hata K, Van Thiel DH, Herberman RB, et al. Phenotypic and functional characteristics of lymphocytes isolated from liver biopsy specimens from patients with active liver disease. Hepatology 1992;15:816–23.
- [60] Cameron RG, Blendis LM, Neuman MG. Accumulation of macrophages in primary sclerosing cholangitis. Clin Biochem 2001;34:195–201.
- [61] Whiteside TL, Lasky S, Si L, et al. Immunologic analysis of mononuclear cells in liver tissues and blood of patients with primary sclerosing cholangitis. Hepatology 1985;5:468–74.
- [62] Ponsioen CY, Kuiper H, Ten Kate FJ, et al. Immunohistochemical analysis of inflammation in primary sclerosing cholangitis. Eur J Gastroenterol Hepatol 1999;11:769–74.
- [63] Adams DH, Eksteen B. Aberrant homing of mucosal T cells and extraintestinal manifestations of inflammatory bowel disease. Nat Rev Immunol 2006;6:244–51.
- [64] Grant AJ, Lalor PF, Salmi M, et al. Homing of mucosal lymphocytes to the liver in the pathogenesis of hepatic complications of inflammatory bowel disease. Lancet 2002;359:150–7.
- [65] Hillan KJ, Hagler KE, MacSween RN, et al. Expression of the mucosal vascular addressin, MAdCAM-1, in inflammatory liver disease. Liver 1999;19: 509–18.
- [66] Grant AJ, Lalor PF, Hubscher SG, et al. MAdCAM-1 expressed in chronic inflammatory liver disease supports mucosal lymphocyte adhesion to hepatic endothelium (MAdCAM-1 in chronic inflammatory liver disease). Hepatology 2001;33:1065–72.
- [67] Eksteen B, Grant AJ, Miles A, et al. Hepatic endothelial CCL25 mediates the recruitment of CCR9+ gut-homing lymphocytes to the liver in primary sclerosing cholangitis. J Exp Med 2004;200:1511–7.
- [68] Chapman RW, Cottone M, Selby WS, et al. Serum autoantibodies, ulcerative colitis and primary sclerosing cholangitis. Gut 1986;27:86–91.
- [69] Mandal A, Dasgupta A, Jeffers L, et al. Autoantibodies in sclerosing cholangitis against a shared peptide in biliary and colon epithelium. Gastroenterology 1994;106:185–92.
- [70] Xu B, Broome U, Ericzon BG, et al. High frequency of autoantibodies in patients with primary sclerosing cholangitis that bind biliary epithelial cells and induce expression of CD44 and production of interleukin 6. Gut 2002;51:120–7.
- [71] Broome U, Grunewald J, Scheynius A, et al. Preferential V beta 3 usage by hepatic T lymphocytes in patients with primary sclerosing cholangitis. J Hepatol 1997;26:527–34.
- [72] Terjung B, Herzog V, Worman HJ, et al. Atypical antineutrophil cytoplasmic antibodies with perinuclear fluorescence in chronic inflammatory bowel diseases and hepatobiliary disorders colocalize with nuclear lamina proteins. Hepatology 1998;28:332–40.
- [73] Terjung B, Spengler U, Sauerbruch T, et al. "Atypical p-ANCA" in IBD and hepatobiliary disorders react with a 50-kilodalton nuclear envelope protein of neutrophils and myeloid cell lines. Gastroenterology 2000;119: 310-22.
- [74] Billing P, Tahir S, Calfin B, et al. Nuclear localization of the antigen detected by ulcerative colitis-associated perinuclear antineutrophil cytoplasmic antibodies. Am J Pathol 1995;147:979–87.
- [75] Klein R, Eisenburg J, Weber P, et al. Significance and specificity of antibodies to neutrophils detected by western blotting for the serological diagnosis of primary sclerosing cholangitis. Hepatology 1991;14:1147–52.
- [76] Terjung B, Soehne J, Lechtenberg B, et al. p-ANCA in autoimmune liver disorders recognize human beta-tubulin isotype 5 and cross-react with microbial protein FtsZ. Gut, doi:10.1136/gut.2008.157818.
- [77] Smit JJ, Schinkel AH, Oude Elferink RP, et al. Homozygous disruption of the murine mdr2 P-glycoprotein gene leads to a complete absence of phospholipid from bile and to liver disease. Cell 1993;75:451–62.
- [78] Trauner M, Fickert P, Wagner M. MDR3 (ABCB4) defects: a paradigm for the genetics of adult cholestatic syndromes. Semin Liver Dis 2007;27:77–98.
- [79] Popov Y, Patsenker E, Fickert P, et al. Mdr2 (Abcb4)-/- mice spontaneously develop severe biliary fibrosis via massive dysregulation of pro- and antifibrogenic genes. J Hepatol 2005;43:1045-54.
- [80] Fickert P, Fuchsbichler A, Wagner M, et al. Regurgitation of bile acids from leaky bile ducts causes sclerosing cholangitis in Mdr2 (Abcb4) knockout mice. Gastroenterology 2004;127:261–74.
- [81] Fickert P, Zollner G, Fuchsbichler A, et al. Ursodeoxycholic acid aggravates bile infarcts in bile duct-ligated and Mdr2 knockout mice via disruption of cholangioles. Gastroenterology 2002;123:1238–51.
- [82] Nakken KE, Nygard S, Haaland T, et al. Multiple inflammatory-, tissue remodelling- and fibrosis genes are differentially transcribed in the livers of Abcb4 (-/-) mice harbouring chronic cholangitis. Scand J Gastroenterol 2007;42:1245-55.
- [83] Jacquemin E. Role of multidrug resistance 3 deficiency in pediatric and adult liver disease: one gene for three diseases. Semin Liver Dis 2001;21:551–62.

- [84] Ziol M, Barbu V, Rosmorduc O, et al. ABCB4 heterozygous gene mutations associated with fibrosing cholestatic liver disease in adults. Gastroenterology 2008;135:131–41.
- [85] Gotthardt D, Runz H, Keitel V, et al. A mutation in the canalicular phospholipid transporter gene, ABCB4, is associated with cholestasis, ductopenia, and cirrhosis in adults. Hepatology 2008;48:1157–66.
- [86] Melum E, Boberg KM, Franke A, et al. Variation in the MDR3 gene influences disease progression in PSC patients and disease susceptibility in epistatic interaction with polymorphism in OST-alpha gene. Hepatology 2007;46:265A.
- [87] Ohishi Y, Nakamura M, lio N, et al. Single-nucleotide polymorphism analysis of the multidrug resistance protein 3 gene for the detection of clinical progression in Japanese patients with primary biliary cirrhosis. Hepatology 2008;48:853–62.
- [88] Palmer RH. Bile acids, liver injury, and liver disease. Arch Intern Med 1972;130:606-17.
- [89] Wagner M, Halilbasic E, Marschall HU, et al. CAR and PXR agonists stimulate hepatic bile acid and bilirubin detoxification and elimination pathways in mice. Hepatology 2005;42:420–30.
- [90] Parks DJ, Blanchard SG, Bledsoe RK, et al. Bile acids: natural ligands for an orphan nuclear receptor. Science 1999;284:1365–8.
- [91] Guo GL, Lambert G, Negishi M, et al. Complementary roles of farnesoid X receptor, pregnane X receptor, and constitutive androstane receptor in protection against bile acid toxicity. J Biol Chem 2003;278:45062–71.
- [92] Staudinger JL, Goodwin B, Jones SA, et al. The nuclear receptor PXR is a lithocholic acid sensor that protects against liver toxicity. Proc Natl Acad Sci USA 2001;98:3369–74.
- [93] Stedman CA, Liddle C, Coulter SA, et al. Nuclear receptors constitutive androstane receptor and pregnane X receptor ameliorate cholestatic liver injury. Proc Natl Acad Sci USA 2005;102:2063–8.
- [94] Saini SP, Mu Y, Gong H, et al. Dual role of orphan nuclear receptor pregnane X receptor in bilirubin detoxification in mice. Hepatology 2005;41: 497–505.
- [95] Xie W, Radominska-Pandya A, Shi Y, et al. An essential role for nuclear receptors SXR/PXR in detoxification of cholestatic bile acids. Proc Natl Acad Sci USA 2001;98:3375–80.
- [96] Karlsen TH, Lie BA, Frey Froslie K, et al. Polymorphisms in the steroid and xenobiotic receptor gene influence survival in primary sclerosing cholangitis. Gastroenterology 2006;131:781–7.
- [97] Sookoian S, Castano GO, Burgueno AL, et al. The nuclear receptor PXR gene variants are associated with liver injury in nonalcoholic fatty liver disease. Pharmacogenet Genomics 2010;20:1–8.
- [98] Poupon R, Ping C, Chretien Y, et al. Genetic factors of susceptibility and of severity in primary biliary cirrhosis. J Hepatol 2008;49:1038–45.
- [99] Lichtman SN, Okoruwa EE, Keku J, et al. Degradation of endogenous bacterial cell wall polymers by the muralytic enzyme mutanolysin prevents hepatobiliary injury in genetically susceptible rats with experimental intestinal bacterial overgrowth. J Clin Invest 1992;90:1313–22.
- [100] Lichtman SN, Keku J, Schwab JH, et al. Evidence for peptidoglycan absorption in rats with experimental small bowel bacterial overgrowth. Infect Immun 1991;59:555–62.
- [101] Lichtman SN, Keku J, Schwab JH, et al. Hepatic injury associated with small bowel bacterial overgrowth in rats is prevented by metronidazole and tetracycline. Gastroenterology 1991;100:513–9.
- [102] Lichtman SN, Keku J, Clark RL, et al. Biliary tract disease in rats with experimental small bowel bacterial overgrowth. Hepatology 1991;13:766–72.
- [103] Lichtman SN, Sartor RB, Keku J, et al. Hepatic inflammation in rats with experimental small intestinal bacterial overgrowth. Gastroenterology 1990;98:414-23.
- [104] Bo X, Broome U, Remberger M, et al. Tumour necrosis factor alpha impairs function of liver derived T lymphocytes and natural killer cells in patients with primary sclerosing cholangitis. Gut 2001;49:131–41.
- [105] Berg PA, Klein R, Rocken M. Cytokines in primary biliary cirrhosis. Semin Liver Dis 1997;17:115–23.
- [106] Aoki CA, Dawson K, Kenny TP, et al. Gene expression by PBMC in primary sclerosing cholangitis: evidence for dysregulation of immune mediated genes. Clin Dev Immunol 2006;13:265–71.
- [107] Karrar A, Broome U, Södergren T, et al. Biliary epithelial cell antibodies link adaptive and innate immune responses in primary sclerosing cholangitis. Gastroenterology 2007;4:1504–14.
- [108] Farkkila M, Karvonen AL, Nurmi H, et al. Metronidazole and ursodeoxycholic acid for primary sclerosing cholangitis: a randomized placebo-controlled trial. Hepatology 2004;40:1379–86.
- [109] Blechacz B, Gores GJ. Cholangiocarcinoma: advances in pathogenesis, diagnosis, and treatment. Hepatology 2008;48:308–21.
- [110] Komichi D, Tazuma S, Nishioka T, et al. Glycochenodeoxycholate plays a carcinogenic role in immortalized mouse cholangiocytes via oxidative DNA damage. Free Radic Biol Med 2005;39:1418–27.
- [111] Forsbring M, Vik ES, Dalhus B, et al. Catalytically impaired hMYH and NEIL1 mutant proteins identified in patients with primary sclerosing cholangitis and cholangiocarcinoma. Carcinogenesis 2009;30:1147–54.
- [112] Kobayashi S, Werneburg NW, Bronk SF, et al. Interleukin-6 contributes to Mcl-1 up-regulation and TRAIL resistance via an Akt-signaling pathway in cholangiocarcinoma cells. Gastroenterology 2005;128:2054–65.
- [113] Bromberg J, Wang TC. Inflammation and cancer: IL-6 and STAT3 complete the link. Cancer Cell 2009;15:79–80.

- [114] Isomoto H, Mott JL, Kobayashi S, et al. Sustained IL-6/STAT-3 signaling in cholangiocarcinoma cells due to SOCS-3 epigenetic silencing. Gastroenterology 2007;132:384–96.
- [115] Isomoto H. Epigenetic alterations associated with cholangiocarcinoma (review). Oncol Rep 2009;22:227–32.
- [116] Condeelis J, Pollard JW. Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. Cell 2006;124:263–6.
- [117] Guerra N, Tan YX, Joncker NT, et al. NKG2D-deficient mice are defective in tumor surveillance in models of spontaneous malignancy. Immunity 2008;28:571–80.
- [118] Melum E, Karlsen TH, Schrumpf E, et al. Cholangiocarcinoma in primary sclerosing cholangitis is associated with NKG2D polymorphisms. Hepatology 2008;47:90–6.
- [119] Hayashi T, Imai K, Morishita Y, et al. Identification of the NKG2D haplotypes associated with natural cytotoxic activity of peripheral blood lymphocytes and cancer immunosurveillance. Cancer Res 2006;66:563–70.
- [120] Boberg KM, Bergquist A, Mitchell S, et al. Cholangiocarcinoma in primary sclerosing cholangitis: risk factors and clinical presentation. Scand J Gastroenterol 2002;37:1205–11.
- [121] Ahrendt SA, Pitt HA, Nakeeb A, et al. Diagnosis and management of cholangiocarcinoma in primary sclerosing cholangitis. J Gastrointest Surg 1999;3:357–67, discussion 67–68.
- [122] MacCarty RL, LaRusso NF, Wiesner RH, et al. Primary sclerosing cholangitis: findings on cholangiography and pancreatography. Radiology 1983;149:39-44.
- [123] EASL clinical practice guidelines: management of cholestatic liver diseases. J Hepatol 2009;51:237–67.
- [124] Chapman RW, Fevery J, Kalloo AN, et al. AASLD guidelines: diagnosis and management of primary sclerosing cholangitis (PSC). Hepatology 2010;51:660–78.
- [125] Angulo P, Pearce DH, Johnson CD, et al. Magnetic resonance cholangiography in patients with biliary disease: its role in primary sclerosing cholangitis. J Hepatol 2000;33:520–7.
- [126] Moff SL, Kamel IR, Eustace J, et al. Diagnosis of primary sclerosing cholangitis: a blinded comparative study using magnetic resonance cholangiography and endoscopic retrograde cholangiography. Gastrointest Endosc 2006;64:219–23.
- [127] Vitellas KM, El-Dieb A, Vaswani KK, et al. MR cholangiopancreatography in patients with primary sclerosing cholangitis: interobserver variability and comparison with endoscopic retrograde cholangiopancreatography. AJR Am J Roentgenol 2002;179:399–407.
- [128] Webster GJ, Pereira SP, Chapman RW. Autoimmune pancreatitis/IgG4associated cholangitis and primary sclerosing cholangitis—overlapping or separate diseases? J Hepatol 2009;51:398–402.
- [129] Bjornsson E. Immunoglobulin G4-associated cholangitis. Curr Opin Gastroenterol 2008;24:389–94.
- [130] Mendes FD, Jorgensen R, Keach J, et al. Elevated serum IgG4 concentration in patients with primary sclerosing cholangitis. Am J Gastroenterol 2006;101:2070–5.
- [131] Bjornsson E, Olsson R, Bergquist A, et al. The natural history of small-duct primary sclerosing cholangitis. Gastroenterology 2008;134:975–80.
- [132] Angulo P, Maor-Kendler Y, Lindor KD. Small-duct primary sclerosing cholangitis: a long-term follow-up study. Hepatology 2002;35:1494–500.
- [133] Boberg KM, Schrumpf E, Fausa O, et al. Hepatobiliary disease in ulcerative colitis. An analysis of 18 patients with hepatobiliary lesions classified as small-duct primary sclerosing cholangitis. Scand J Gastroenterol 1994;29: 744–52.
- [134] Broome U, Glaumann H, Lindstom E, et al. Natural history and outcome in 32 Swedish patients with small duct primary sclerosing cholangitis (PSC). J Hepatol 2002;36:586–9.
- [135] Rabinovitz M, Demetris AJ, Bou-Abboud CF, et al. Simultaneous occurrence of primary sclerosing cholangitis and autoimmune chronic active hepatitis in a patient with ulcerative colitis. Dig Dis Sci 1992;37:1606–11.
- [136] Lawrence SP, Sherman KE, Lawson JM, et al. A 39 year old man with chronic hepatitis. Semin Liver Dis 1994;14:97–105.
- [137] Perdigoto R, Carpenter HA, Czaja AJ. Frequency and significance of chronic ulcerative colitis in severe corticosteroid-treated autoimmune hepatitis. J Hepatol 1992;14:325–31.
- [138] Floreani A, Rizzotto ER, Ferrara F, et al. Clinical course and outcome of autoimmune hepatitis/primary sclerosing cholangitis overlap syndrome. Am J Gastroenterol 2005;100:1516–22.
- [139] Debray D, Pariente D, Urvoas E, et al. Sclerosing cholangitis in children. J Pediatr 1994;124:49–56.
- [140] Wilschanski M, Chait P, Wade JA, et al. Primary sclerosing cholangitis in 32 children: clinical, laboratory, and radiographic features, with survival analysis. Hepatology 1995;22:1415–22.
- [141] Feldstein AE, Perrault J, El-Youssif M, et al. Primary sclerosing cholangitis in children: a long-term follow-up study. Hepatology 2003;38:210–7.
- [142] Batres LA, Russo P, Mathews M, et al. Primary sclerosing cholangitis in children: a histologic follow-up study. Pediatr Dev Pathol 2005;8:568–76.
- [143] Miloh T, Arnon R, Shneider B, et al. A retrospective single-center review of primary sclerosing cholangitis in children. Clin Gastroenterol Hepatol 2009;7:239–45.
- [144] Gregorio GV, Portmann B, Karani J, et al. Autoimmune hepatitis/sclerosing cholangitis overlap syndrome in childhood: a 16-year prospective study. Hepatology 2001;33:544–53.

- [145] McNair AN, Moloney M, Portmann BC, et al. Autoimmune hepatitis overlapping with primary sclerosing cholangitis in five cases. Am J Gastroenterol 1998;93:777–84.
- [146] Boberg KM, Egeland T, Schrumpf E. Long-term effect of corticosteroid treatment in primary sclerosing cholangitis patients. Scand J Gastroenterol 2003;38:991–5.
- [147] Rosen CB, Nagorney DM, Wiesner RH, et al. Cholangiocarcinoma complicating primary sclerosing cholangitis. Ann Surg 1991;213:21–5.
- [148] Bergquist A, Glaumann H, Persson B, et al. Risk factors and clinical presentation of hepatobiliary carcinoma in patients with primary sclerosing cholangitis: a case-control study. Hepatology 1998;27:311-6.
- [149] Chalasani N, Baluyut A, Ismail A, et al. Cholangiocarcinoma in patients with primary sclerosing cholangitis: a multicenter case-control study. Hepatology 2000;31:7-11.
- [150] Blechacz BR, Sanchez W, Gores GJ. A conceptual proposal for staging ductal cholangiocarcinoma. Curr Opin Gastroenterol 2009;25:238–9.
- [151] Khan SA, Davidson BR, Goldin R, et al. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. Gut 2002;51:VI1-9.
- [152] Malhi H, Gores GJ. Review article: the modern diagnosis and therapy of cholangiocarcinoma. Aliment Pharmacol Ther 2006;23:1287–96.
- [153] Campbell WL, Ferris JV, Holbert BL, et al. Biliary tract carcinoma complicating primary sclerosing cholangitis: evaluation with CT, cholangiography, US, and MR imaging. Radiology 1998;207:41–50.
- [154] Teefey SA, Baron RL, Rohrmann CA, et al. Sclerosing cholangitis: CT findings. Radiology 1988;169:635–9.
- [155] Zhang Y, Uchida M, Abe T, et al. Intrahepatic peripheral cholangiocarcinoma: comparison of dynamic CT and dynamic MRI. J Comput Assist Tomogr 1999;23:670-7.
- [156] Lee MG, Park KB, Shin YM, et al. Preoperative evaluation of hilar cholangiocarcinoma with contrast-enhanced three-dimensional fast imaging with steady-state precession magnetic resonance angiography: comparison with intraarterial digital subtraction angiography. World J Surg 2003;27: 278–83.
- [157] Kluge R, Schmidt F, Caca K, et al. Positron emission tomography with [(18)F]fluoro-2-deoxy-D-glucose for diagnosis and staging of bile duct cancer. Hepatology 2001;33:1029–35.
- [158] Prytz H, Keiding S, Bjornsson E, et al. Dynamic FDG-PET is useful for detection of cholangiocarcinoma in patients with PSC listed for liver transplantation. Hepatology 2006;44:1572–80.
- [159] Anderson CD, Rice MH, Pinson CW, et al. Fluorodeoxyglucose PET imaging in the evaluation of gallbladder carcinoma and cholangiocarcinoma. J Gastrointest Surg 2004;8:90–7.
- [160] Fevery J, Buchel O, Nevens F, et al. Positron emission tomography is not a reliable method for the early diagnosis of cholangiocarcinoma in patients with primary sclerosing cholangitis. J Hepatol 2005;43:358–60.
- [161] Tischendorf JJ, Kruger M, Trautwein Č, et al. Cholangioscopic characterization of dominant bile duct stenoses in patients with primary sclerosing cholangitis. Endoscopy 2006;38:665–9.
- [162] Tischendorf JJ, Meier PN, Schneider A, et al. Transpapillary intraductal ultrasound in the evaluation of dominant bile duct stenoses in patients with primary sclerosing cholangitis. Scand | Gastroenterol 2007;42:1011–7.
- [163] Charatcharoenwitthaya P, Enders FB, Halling KC, et al. Utility of serum tumor markers, imaging, and biliary cytology for detecting cholangiocarcinoma in primary sclerosing cholangitis. Hepatology 2008;48:1106–17.
- [164] Moreno Luna LE, Kipp B, Halling KC, et al. Advanced cytologic techniques for the detection of malignant pancreatobiliary strictures. Gastroenterology 2006;131:1064–72.
- [165] Kipp BR, Stadheim LM, Halling SA, et al. A comparison of routine cytology and fluorescence in situ hybridization for the detection of malignant bile duct strictures. Am J Gastroenterol 2004;99:1675–81.
- [166] Baron TH, Harewood GC, Rumalla A, et al. A prospective comparison of digital image analysis and routine cytology for the identification of malignancy in biliary tract strictures. Clin Gastroenterol Hepatol 2004;2:214–9.
- [167] Bonney GK, Craven RA, Prasad R, et al. Circulating markers of biliary malignancy: opportunities in proteomics? Lancet Oncol 2008;9:149–58.
- [168] Mullighan CG, Goorha S, Radtke I, et al. Genome-wide analysis of genetic alterations in acute lymphoblastic leukaemia. Nature 2007;446:758–64.
- [169] Miller G, Socci ND, Dhall D, et al. Genome wide analysis and clinical correlation of chromosomal and transcriptional mutations in cancers of the biliary tract. J Exp Clin Cancer Res 2009;28:62.
- [170] Spratlin JL, Serkova NJ, Eckhardt SG. Clinical applications of metabolomics in oncology: a review. Clin Cancer Res 2009;15:431–40.
- [171] Alvaro D. Serum and bile biomarkers for cholangiocarcinoma. Curr Opin Gastroenterol 2009;25:279–84.
- [172] Alvaro D, Macarri G, Mancino MG, et al. Serum and biliary insulin-like growth factor I and vascular endothelial growth factor in determining the cause of obstructive cholestasis. Ann Intern Med 2007;147:451–9.
- [173] Stiehl A, Walker S, Stiehl L, et al. Effect of ursodeoxycholic acid on liver and bile duct disease in primary sclerosing cholangitis. A 3-year pilot study with a placebo-controlled study period. J Hepatol 1994;20:57–64.
- [174] Mitchell SA, Bansi DS, Hunt N, et al. A preliminary trial of high-dose ursodeoxycholic acid in primary sclerosing cholangitis. Gastroenterology 2001;121:900–7.
- [175] Beuers U, Spengler U, Kruis W, et al. Ursodeoxycholic acid for treatment of primary sclerosing cholangitis: a placebo-controlled trial. Hepatology 1992;16:707–14.

- [176] Lindor KD. Ursodiol for primary sclerosing cholangitis. Mayo Primary Sclerosing Cholangitis-Ursodeoxycholic Acid Study Group. N Engl J Med 1997;336:691–5.
- [177] Olsson R, Boberg KM, de Muckadell OS, et al. High-dose ursodeoxycholic acid in primary sclerosing cholangitis: a 5-year multicenter, randomized, controlled study. Gastroenterology 2005;129:1464–72.
- [178] Lindor KD, Kowdley KV, Luketic VA, et al. High-dose ursodeoxycholic acid for the treatment of primary sclerosing cholangitis. Hepatology 2009;50: 808–14.
- [179] Shi J, Li Z, Zeng X, et al. Ursodeoxycholic acid in primary sclerosing cholangitis: meta-analysis of randomized controlled trials. Hepatol Res 2009;39: 865–73.
- [180] Cullen SN, Rust C, Fleming K, et al. High dose ursodeoxycholic acid for the treatment of primary sclerosing cholangitis is safe and effective. J Hepatol 2008;48:792–800.
- [181] Harnois DM, Angulo P, Jorgensen RA, et al. High-dose ursodeoxycholic acid as a therapy for patients with primary sclerosing cholangitis. Am J Gastroenterol 2001;96:1558–62.
- [182] Rudolph G, Kloeters-Plachky P, Rost D, et al. The incidence of cholangiocarcinoma in primary sclerosing cholangitis after long-time treatment with ursodeoxycholic acid. Eur J Gastroenterol Hepatol 2007;19:487–91.
- [183] Brandsaeter B, Isoniemi H, Broome U, et al. Liver transplantation for primary sclerosing cholangitis; predictors and consequences of hepatobiliary malignancy. J Hepatol 2004;40:815–22.
- [184] Tung BY, Emond MJ, Haggitt RC, et al. Ursodiol use is associated with lower prevalence of colonic neoplasia in patients with ulcerative colitis and primary sclerosing cholangitis. Ann Intern Med 2001;134:89–95.
- [185] Pardi DS, Loftus Jr EV, Kremers WK, et al. Ursodeoxycholic acid as a chemopreventive agent in patients with ulcerative colitis and primary sclerosing cholangitis. Gastroenterology 2003;124:889–93.
- [186] Sjoqvist U, Tribukait B, Ost A, et al. Ursodeoxycholic acid treatment in IBD-patients with colorectal dysplasia and/or DNA-aneuploidy: a prospective, double-blind, randomized controlled pilot study. Anticancer Res 2004;24:3121–7.
- [187] Cullen SN, Chapman RW. The medical management of primary sclerosing cholangitis. Semin Liver Dis 2006;26:52–61.
- [188] Luth S, Kanzler S, Frenzel C, et al. Characteristics and long-term prognosis of the autoimmune hepatitis/primary sclerosing cholangitis overlap syndrome. J Clin Gastroenterol; in press.
- [189] Ghazale A, Chari ST, Zhang L, et al. Immunoglobulin G4-associated cholangitis:
- clinical profile and response to therapy. Gastroenterology 2008;134:706–15. [190] Church NI, Pereira SP, Deheragoda MG, et al. Autoimmune pancreatitis: clinical and radiological features and objective response to steroid therapy in a
- UK series. Am J Gastroenterol 2007;102:2417–25. [191] Hirano K, Tada M, Isayama H, et al. Long-term prognosis of autoimmune pancreatitis with and without corticosteroid treatment. Gut 2007:56:1719–24.
- [192] Stiehl A, Rudolph G, Kloters-Plachky P, et al. Development of dominant bile duct stenoses in patients with primary sclerosing cholangitis treated with ursodeoxycholic acid: outcome after endoscopic treatment. J Hepatol 2002;36:151–6.
- [193] Baluyut AR, Sherman S, Lehman GA, et al. Impact of endoscopic therapy on the survival of patients with primary sclerosing cholangitis. Gastrointest Endosc 2001;53:308–12.
- [194] Gluck M, Cantone NR, Brandabur JJ, et al. A twenty-year experience with endoscopic therapy for symptomatic primary sclerosing cholangitis. J Clin Gastroenterol 2008;42:1032–9.
- [195] Kaya M, Petersen BT, Angulo P, et al. Balloon dilation compared to stenting of dominant strictures in primary sclerosing cholangitis. Am J Gastroenterol 2001;96:1059–66.
- [196] Stiehl A. Primary sclerosing cholangitis: the role of endoscopic therapy. Semin Liver Dis 2006;26:62–8.
- [197] Ponsioen CY, Lam K, van Milligen de Wit AW, et al. Four years experience with short term stenting in primary sclerosing cholangitis. Am J Gastroenterol 1999;94:2403–7.
- [198] van Milligen de Wit AW, Rauws EA, van Bracht J, et al. Lack of complications following short-term stent therapy for extrahepatic bile duct strictures in primary sclerosing cholangitis. Gastrointest Endosc 1997;46:344–7.
- [199] Bjoro K, Brandsaeter B, Foss A, et al. Liver transplantation in primary sclerosing cholangitis. Semin Liver Dis 2006;26:69–79.
- [200] Bjoro K, Schrumpf E. Liver transplantation for primary sclerosing cholangitis. J Hepatol 2004;40:570–7.
- [201] Uemura T, Ikegami T, Sanchez EQ, et al. Late acute rejection after liver transplantation impacts patient survival. Clin Transplant 2008;22:316–23.
- [202] Graziadei IW, Wiesner RH, Marotta PJ, et al. Long-term results of patients undergoing liver transplantation for primary sclerosing cholangitis. Hepatology 1999;30:1121–7.
- [203] Neuberger J. Incidence, timing, and risk factors for acute and chronic rejection. Liver Transpl Surg 1999;5:S30–6.
- [204] Narumi S, Roberts JP, Emond JC, et al. Liver transplantation for sclerosing cholangitis. Hepatology 1995;22:451–7.
- [205] Shaked A, Colonna JO, Goldstein L, et al. The interrelation between sclerosing cholangitis and ulcerative colitis in patients undergoing liver transplantation. Ann Surg 1992;215:598–603.
- [206] Miki C, Harrison JD, Gunson BK, et al. Inflammatory bowel disease in primary sclerosing cholangitis: an analysis of patients undergoing liver transplantation. Br J Surg 1995;82:1114–7.

- [207] Kugelmas M, Spiegelman P, Osgood MJ, et al. Different immunosuppressive regimens and recurrence of primary sclerosing cholangitis after liver transplantation. Liver Transpl 2003;9:727–32.
- [208] Hoffman CEE. Verschluss der Gallenwege durch Verdickung der Wandungen. Arch Pathol Anat Physiol 1867;49:206–15.
- [209] Smith MP, Loe RH. Sclerosing cholangitis; review of recent case reports and associated diseases and four new cases. Am J Surg 1965;110:239–46.
- [210] Warren KW, Athanassiades S. Monge JI Primary sclerosing cholangitis. A study of forty-two cases. Am J Surg 1966;111:23–38.
- [211] McFarlane IG, Wojcicka BM, Tsantoulas DC, et al. Leukocyte migration inhibition in response to biliary antigens in primary biliary cirrhosis, sclerosing cholangitis, and other chronic liver diseases. Gastroenterology 1979;76:1333–40.
- [212] Chapman RW, Arborgh BA, Rhodes JM, et al. Primary sclerosing cholangitis: a review of its clinical features, cholangiography, and hepatic histology. Gut 1980;21:870–7.
- [213] Schrumpf E, Elgjo K, Fausa O, et al. Sclerosing cholangitis in ulcerative colitis. Scand J Gastroenterol 1980;15:689–97.
- [214] NIH National Institutes of Health Consensus Development Conference Statement: liver transplantation—June 20–23, 1983. Hepatology 1984:4:107S-10S.
- [215] Wee A, Ludwig J, Coffey Jr RJ, et al. Hepatobiliary carcinoma associated with primary sclerosing cholangitis and chronic ulcerative colitis. Hum Pathol 1985;16:719–26.

- [216] Wee A, Ludwig J. Pericholangitis in chronic ulcerative colitis: primary sclerosing cholangitis of the small bile ducts? Ann Intern Med 1985;102: 581-7.
- [217] Lerut J, Demetris AJ, Stieber AC, et al. Intrahepatic bile duct strictures after human orthotopic liver transplantation. Recurrence of primary sclerosing cholangitis or unusual presentation of allograft rejection? Transpl Int 1988;1:127–30.
- [218] Broome U, Lindberg G, Lofberg R. Primary sclerosing cholangitis in ulcerative colitis—a risk factor for the development of dysplasia and DNA aneuploidy? Gastroenterology 1992;102:1877–80.
- [219] Bergquist A, Lindberg G, Saarinen S, et al. Increased prevalence of primary sclerosing cholangitis among first-degree relatives. J Hepatol 2005;42: 252–6.
- [220] Hov JR, Boberg KM, Karlsen TH. Autoantibodies in primary sclerosing cholangitis. World J Gastroenterol 2008;14:3781–91.
- [221] Ueno Y, Phillips JO, Ludwig J, et al. Development and characterization of a rodent model of immune-mediated cholangitis. Proc Natl Acad Sci USA 1996;93:216–20.
- [222] Boberg KM, Fausa O, Haaland T, et al. Features of autoimmune hepatitis in primary sclerosing cholangitis: an evaluation of 114 primary sclerosing cholangitis patients according to a scoring system for the diagnosis of autoimmune hepatitis. Hepatology 1996;23:1369–76.
- [223] Colombo C. Liver disease in cystic fibrosis. Curr Opin Pulm Med 2007;13:529–36.